Study of antimicrobial effect of some plants of Lamiaceae family on *Escherichia coli* species isolated from children with urinary tract infection

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INTRODUCTION

Urinary tract infection (UTI) is a common disease in children. About 1%–3% of girls and 1% of boys suffer from UTI. Gram-negative bacteria in intestine are the main cause of UTI. *Escherichia coli* is the most common bacteria in UTI, causing 75%–90% of all infections, followed by *Klebsiella* and *Proteus* (1). The majority of UTI cases are ascending infections. The bacteria originated from stool flora are colonized in perineal area and reach the bladder through urethra. The management of UTI
patients is faced with microbial resistance. Antibiotics currently used in the treatment of UTI gradually lose their effects. Although some antibiotics, such as co-trimoxazole, ampicillin, and amoxicillin are inexpensive, the administration of them is not suitable in many countries because of their ineffectiveness (2). Risk factors, such as genitourinary system anomalies, vesicoureteral reflux, history of antibiotic administration, and recurrent UTI are important factors in increasing the incidence of antibiotic resistance. Unnecessary consumption of antibiotics results in the bacterial resistance to available and commonly used drugs, which in turn leads to increase the administration of newer and more expensive drugs. Continuing this trend results in bacterial resistance to newer drugs.

Medicinal plants and herbal medicine have been one of the most important issues in medical and pharmacological sciences. The use of medicinal plants and natural products has a long history. Following numerous failures in the use of chemical drugs and the advent of new resistant microorganisms, human beings have been producing various medicinal products with herbal origin. Lamiaceae family has more than 2500 members and mostly grows in temperate regions. They are indigenous to the Mediterranean region. Eighty-one species of Lamiaceae family are used for medicinal purposes in Iran. They have been used in traditional treatment of bloating and indigestion, insomnia, psychological disorders such as depression, anxiety and for memory improvement (3). The main compounds in the Lamiaceae family plants are flavonoids and polyphenols. These compounds are inhibitors of cholinesterase and their antioxidant and anti-inflammatory effects have been reported in several studies (4-6). In the previous studies, the antimicrobial effects of the essential oils of some members of Lamiaceae family, such as thyme, savory, marjoram, basil and lavender have been investigated (7-9).

To the best of our knowledge, there is no published study investigating the antibacterial effects of medicinal plants on E. coli, the most common pathogen of UTI. Therefore, the present study was performed to investigate the antibacterial effects of 13 species of Lamiaceae family, including Salvia spinosa, Salvia multicaulis, Salvia virgata, Stachys lavandulifolia, Stachys inflata, Nepeta crispa, Nepeta fissa, Phlomis olivieri, Phlomis auberti, Phlomis herba-venti, Phlomis anisodontea, Thymus daenensis, and Teucrium polium against E. coli, and compared with antibacterial effects of the common antibiotics used in UTI, including cefixime, nitrofurantoin, co-trimoxazole, amikacin, and ceftriaxone.

Materials and Methods
The plant species were collected from the populations growing wild in Hamadan province, west of Iran. A voucher number was deposited to each specimen in herbarium of Department of Pharmacognosy, School of Pharmacy, Hamadan University of Medical Sciences, Hamadan, Iran. The hydroalcoholic extracts of 13 plant species from Lamiaceae family were prepared and their antimicrobial effects against E. coli were investigated. The air-dried plant materials were powdered and macerated in hydroalcohol (85%) for three times, each time three days. The filtrates were dried by means of rotary evaporator. The obtained extracts were kept in a refrigerator until use. Bacteria isolated from urine samples of children with UTI admitted in the pediatric ward of Besat hospital were studied. After identification, the sensitivity and resistance of the bacteria to the studied extracts and five commonly used antibiotics in the treatment of UTI, including cefixime, nitrofurantoin, co-trimoxazole, amikacin, and ceftriaxone were assessed.

Agar well diffusion assay
To assess the antibacterial activities of the studied extracts, the agar well diffusion method was used. The Mueller-Hinton agar (MHA, Merck Co, Germany) was used according to a previously published method (10) and results were interpreted according to the Clinical & Laboratory Standards Institute (CLSI) guidelines (11). The bacteria were dissolved in Phosphate buffer and their turbidity was compared with 0.5 McFarland standards (10⁶ microorganisms per mL). Agar surface of each plate was streaked with a sterile cotton swab with the bacterial strains. Agar plate was punched with a sterile cork borer of 4 mm size and 50 μL of the 8 different dilutions (from 0.4 mg/mL to 13.5 mg/mL) of the extract was poured with a micropipette in the bore. In each culture plate, one well was considered as a blank. The plates were allowed to stand by for 30 minutes. The plates were incubated at 37°C for 24 hours.

Determination of MIC and MBC
To determine minimum inhibitory concentration (MIC) of each extract, 10 tubes containing different dilutions of the extracts as well as positive and negative controls were used. Different dilutions of each extract from 0.4 mg/mL (tube 1) to 13.5 mg/mL (tube 8) were prepared in nutrient broth culture media (Merck Co., Germany) with one mL of bacteria suspension. The negative control contained 9 mL of the culture medium and one mL of the bacterial suspension. The positive control contained 9 mL of the culture medium with 1 mL of each extract. All tubes were incubated at 37°C for 24 hours and then assessed for their turbidity caused by the growth of inoculated bacteria. For determination of minimum bactericidal concentration (MBC), a sample was taken from all tubes where the bacterium was inhibited in the MIC assay. After the cultivation of MHA and 24 hours incubation, the results were studied.

Ethical issues
The study was conducted in accordance with the principles of Declaration of Helsinki 1996 version and its later
amendments and also Good Clinical Practice standards.
The study protocol and other study related documents were reviewed and approved by research council of Hamadan University of Medical Sciences (# 9211153891).

Statistical analysis
Data were analyzed using SPSS version 23.0. For descriptive statistics mean ± SD was used. After checking for variables normality because of abnormal distribution, Kruskal-Wallis H test was used. For comparison of groups, post hoc test (Bonferroni) was used and P<0.5 was considered as significant.

Results
Comparison of the diameter of zone of inhibition
Figure 1 shows the zone of inhibition (ZOI) for the studied extracts and antibiotics. Among antibiotics amikacin had the largest mean diameter of ZOI and cotrimoxazole had the lowest. The difference of ZOI of amikacin was significant with all antibiotics except nitrofurantoin and ceftriaxone. Zone of inhibition of cotrimoxazole was significantly smaller than all other antibiotics.

Nepeta crispa had the largest mean diameter of ZOI among herbal extracts and S. spinosa had the smallest ZOI. This difference of ZOI of N. crispa was significant only with S. spinosa and S. multicaulis.
The mean diameter of ZOI for amikacin was larger than those of the studied extracts and other antibiotics. There was a significant difference with ZOI of all antibiotics and extracts except of ZOI of N. crispa, P. olivieri, T. polium, nitrofurantoin and ceftriaxone.

ZOI for nitrofurantoin was statistically different from the extracts of S. spinosa, S. multicaulis and cotrimoxazole (P<0.05). In addition, the ZOI for N. crispa and P. olivieri were significantly larger than those of cotrimoxazole (P<0.05). Interestingly ZOI of cefixime was not significantly different from that of all herbal extracts (P>0.05). Co-trimoxazole showed the smallest ZOI among all studied materials and it was significantly different from all studied antibiotic and extracts, except for S. inflata, S. multicaulis and S. spinosa (P<0.05).

MIC and MBC
Figures 2 and 3 show MBC and MIC of studied herbal extracts, respectively. The MIC for N. crispa was significantly lower than other extracts (P<0.05). The MIC for P. olivieri was significantly lower than MIC of S. spinosa, P. aucheri, P. herba-venti, T. daenensis and T. polium (P<0.05). The highest MIC was observed for the extracts of T. polium, which was not significantly different from all other extracts except N. crispa and P. olivieri.

The lowest values of MBC were observed for N. crispa which was significantly lower than those of other extracts (P<0.05) except P. olivieri. T. polium extract had highest MBC but was not significantly different from other extracts except N. crispa and P. olivieri. After N. crispa and P. olivieri, S. multicaulis had rather low MBC, but that was not significantly different with other extracts.

About 23.8% and 9.52% of the isolated bacteria were resistant to S. multicaulis and S. spinosa extracts, respectively. None of the isolated E. coli were resistant.

Figure 1. Comparison of mean zone of inhibition (ZOI) between different herbal extracts and antibiotics.

Figure 2. Comparison of minimum bactericidal concentration (MBC) between different herbal extracts and antibiotics.

Figure 3. Comparison of minimum inhibitory concentration (MIC) between different herbal extracts and antibiotics.
to other studied extracts. Among isolated *E. coli*, 80.95%, 33.33%, 28.57%, and 9.52% were resistant to co-trimoxazole, ceftriaxone, cefixime, and nitrofurantoin, respectively. None of the isolated bacteria were resistant to amikacin.

**Discussion**

UTI is a common disease in children which is mainly caused by intestinal bacteria (*E. coli*) (1). The early detection and prevention of the disease is important in children and results in the prevention of kidney damages in them (12). Antibiotics that have commonly been used in the treatment of UTI, despite of low cost are not suitable in many countries because of ineffectiveness (13). The unnecessary consumption of antibiotics results in the bacterial resistance to available and commonly used drugs. Therefore, given the advantages of herbal products over chemical and synthetic drugs, such as lower cost and fewer side effects, better antimicrobial activity, and more popularity in society, the use of them will be of a great importance in solving this problem. Because of their therapeutical effects, Lamiaceae family has been used from ancient time to cure different diseases. Several studies showed the presence of polyphenols, flavonoids, and terpenoids in the extracts of different members of this family (14). The antimicrobial, antifungal, antiviral, anti-inflammation, and anticancer effects of some members of Lamiaceae family have been reported (15-17). In this study, the antibacterial effects of 13 species of Lamiaceae family were compared with commonly used antibiotics in UTI. The most effective antibiotic was amikacin. No bacterial resistance was observed for this antibiotic and its ZOI diameter was significantly larger than other studied antibiotics. In accordance with our findings, Tarhani et al reported that *E. coli* resistance to amikacin was 2.4% and the mean resistance of 3.1% was reported for all studied microorganisms (2). Co-trimoxazole acts by inhibiting bacterial synthesis of dihydrofolic acid and blocking production of tetrahydrofolic acid, so blocks two consecutive steps in the biosynthesis of nucleic acids and proteins essential to bacteria. In the present study, co-trimoxazole was found to have the least antibacterial effect so that its ZOI diameter was significantly smaller than other studied antibiotics and most of herbal extracts. The bacterial resistance to this antibiotic was 80.95%. In the study of Tarhani et al, the highest bacterial resistance was found for ampicillin (88.25%) and amoxicillin (86.6%), followed by co-trimoxazole (75.6%). The bacterial resistance to this antibiotic in the cultivated *E. coli* samples was 54.3% (2), which is in accordance with our findings. In this study, the bacterial resistance to cefixime, nitrofurantoin, and ceftriaxone were 28.57%, 9.52%, and 33.33%, respectively and their ZOI diameters were 14.0 mm, 16.86 mm, and 17.41 mm, respectively. Cefixime and ceftriaxone belong to the third cephalosporin antibiotics which inhibit the mucopentide generation and prevent the formation of cell wall leading to osmotic instability and death of the bacteria. Nitrofurantoin is a macrolide, which seems that inhibits 50S ribosomal subunit and thus results in inhibition of protein synthesis and bacteria death. In the study of Tarhain et al, the bacterial resistances to cefixime, nitrofurantoin, and ceftriaxone were 4.7%, 11.8%, and 3.1%, respectively (2). The bacterial resistance to cefixime and ceftriaxone in the present study was higher than the study by Tarhani et al. This finding can be explained, at least in part, by the recent consumption of these antibiotics in most patients whose urine samples were collected. In the study of Prado et al, the highest bacterial resistance was observed for co-trimoxazole (52%) and ampicillin, respectively (18). In the study of Sahm et al, 92.8% and 7.7% of *E. coli* isolations were resistant to co-trimoxazole and nitrofurantoin, respectively (19). Furthermore, Ryu et al reported that the most suitable antibiotic for the treatment of UTI caused by gram-negative bacteria was amikacin (20). Similar to our findings, In the study by Amiri et al, the bacterial resistance to co-trimoxazole, cefixime, nitrofurantoin, ceftriaxone, and amikacin in *E. coli* isolations from the urine samples of subjects with UTI were 80.4%, 97.9%, 9.50%, 95.9%, and 17.5%, respectively (21).

In the present study, the antimicrobial effects of 13 species of Lamiaceae family were investigated. Our findings showed that *N. crispa* had the highest antibacterial effects. Its ZOI diameters was 16.82 mm which was significantly lower than that of amikacin. However, it was not significantly different from the ZOI diameters for ceftriaxone and nitrofurantoin. The ZOI diameters for *N. crispa* and *P. olivieri* extracts were significantly larger than that of co-trimoxazole. The MIC and MBC of *N. crispa* extract were significantly lower than other studied extracts. In addition, no resistance of *E. coli* to these extracts was observed. *Nepeta crispa* is a member of Lamiaceae family, which has been used as sedative, anti-flatulence, antispasm, anticough, antiasthma, aseptic agent, and diuretic (22). The effective ingredients in these plants are 1,8-cineole (62.8%), 4aa-7a-7aa-nepetalactone (10.3%) and 4aa-7a-7aβ-nepetalactone (9.2%) (23). In accordance with our observations, Sonboli et al found that the ZOI of *E. coli* was 16 mm for *N. crispa* and its inhibitory effects were observed even at 1:16 dilution. Sonboli et al also found that gram-positive bacteria, such as *Staphylococcus aureus* and *Bacillus subtilis* were the most sensitive species to the essential oil of *N. crispa* (24). Likewise, Mahboubi et al reported that the inhibitory and bactericidal effects of *Nepeta crispa* on *E. coli* were higher than those of vancomycin, amphothricine B, and gentamicin (25). *Phlomis* represents 70 species in Iran, of which 8 species are exclusive to Iran. Different species of *Phlomis* have been used in the treatment of diabetes, stomach ulcer, hemorrhoids and inflammations (26). In addition, the antibacterial, anticancer, and antioxidant activities have been reported for them (27,28). The main compounds of the essential oil of *P. olivieri*, obtained from its leaves, were
normal-octane (28.77%), camphor (13.32%), 1,8-cineol (12.24%), α-pinene (7.8%), and germacrene D (6.57%) (29). In accordance with our findings, Turkert et al, in a study of antricancer and antibacterial effects of some species of Phlomis showed that P. olivieri had significant antibacterial effects against E. coli, S. aureus, Klebsiella pneumonia, Pseudomonas aeruginosa, and S. aeruginosa (30).

In our study, the extracts of N. crispa, P. olivieri, T. daenensis, S. lavandulifolia, P. herba-venti, had relatively high antibacterial effects and showed higher ZOI in comparison with cefixime although there was no significant difference. However, their ZOI was significantly larger than that of co-trimoxazole. Salvia virgata, S. inflata, S. multicaulis, P. anisodonta and S. spinosa showed weak antimicrobial effects and their ZOI was smaller than that of cefixime. However, the difference was not significant for any of these extracts. Although it seems that overall inhibitory effects of herbal extracts may be lower than antibiotics, our findings showed that the proportion of resistant isolates to herbal extracts (2.57%) were significantly lower than those resistant to antibiotics (30.47%). Therefore, with increasing the dose of herbal extracts, it is possible to have remarkable inhibitory effects (similar to antibiotics) against bacteria with fewer side effects than chemical and synthetic antibiotics. Bazzaz et al found that the extracts of S. spinosa, S. virgata, and T. polium had no effects on the growth of E. coli (15). Mojab et al showed that the extract of T. daenensis had inhibitory effects against gram-positive bacteria such as S. aureus, Micrococcus luteus, Enterococcus faecalis, and Streptococcus pyogenes, but they did not observe any inhibitory effects against gram-negative bacteria, including E. coli (31).

Since the plants of Lamiaceae family contain flavonoids, it is noted that most of the antibacterial activity of them is because of the formation of complexes between flavonoids and extracellular proteins, soluble proteins, and cell wall proteins. In addition, hydrophobic flavonoids have the ability to destruct the bacteria plasma membrane (32,33). Therefore, antibacterial effects of these plants may be attributed to their different amounts of flavonoids. Thus, the isolation and purification of effective antibacterial compounds in these plants have a great importance in the treatment of UTIs.

Conclusion
Because of the high bacterial resistance to co-trimoxazole, its use in the early treatment of UTIs in our population is not recommended. On the other hand, amikacin and nitrofurantoin showed higher effectiveness than other studied antibiotics and thus they are recommended to be used as suitable antibiotics in the early treatment of UTIs. Considering the important fact that nitrofurantoin has no significant parenchymal concentration in kidney and is not recommended for treatment of pyelonephritis. The studied herbal extracts showed acceptable antibacterial effects. Therefore, because of the high cost and side effects of chemical and synthesized antibiotics, the use of herbal extracts, especially N. crispa may be recommended as an alternative treatment of UTI.

Limitations of our study
One of limitations of our study was that we were unable to exactly specify strains of E. coli isolated from our patients and study their sensitivity and resistance to antibiotics and herbal extracts. Another limitation was that we only determined MIC and MBC of herbal extracts and not antibiotics due to some microbiologic technical problems.

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Authors’ contribution
HEM conceived the study. MYA performed the experiments. MA collected samples. FE analyzed the data. HEM and ShM drafted the final manuscript. All authors read, revised and approved the final manuscript.

Conflicts of interest
The authors declare no potential conflicts of interest.

Ethical considerations
Ethical issues (including plagiarism, data fabrication, double publication) have been completely observed by the authors.

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References
N. crispa, N. rivularis methanolic extract, Benth, from north of Iran. Daru.


